PHYTOCHEMICAL EVALUATION AND IDENTIFICATION OF BIOACTIVE COMPOUNDS IN CAMEL THORN *ALHAGI PSEUDALHAGI* (M. BIEB.) DESV. EX B. KELLER & SHAP. STEM

Wagay N.A.*¹, Yatoo Ghulam Mohiuddin², Khan N.A.³

¹Botany Research Laboratory, Vidyabharati Mahavidyalya College, Amravati, M.S., (India)
²Department of Botany, Govt. Degree College, Beerwah, Budgam, J & K, (India)
³Department of Botany, Faculty of Life sciences, Punjabi University, Patiala, Punjab, (India)

ABSTRACT

Alhagi pseudalhagi (M. Bieb.) Desv. ex B. Keller & Shap. is rarely found in India. It has been reported by various authors for its potential to cure various ailments. In the present study stem of this plant has been analyzed for its chemical constituents. Analytical, Qualitative, Quantitative and GC-MS analysis was carried out for the understudy plant part. Solvents with different polarities were used for the extraction of chemical constituents and those extracts were analyzed by Gas Chromatography- Mass Spectrometric analysis. 17, 9 and 7 compounds were identified in solvents Chloroform, Dichloromethane and 50 % Ethanol respectively among which seven are secondary metabolites. All the seven secondary metabolites are effective against various diseases and have vast pharmacological importance.

Keywords: Medicinal plants, GC-MS, Bioactive compounds

I. INTRODUCTION

Basic medicinal and pharmacological properties of plants are often related to phytochemicals or bioactive compounds. Phytochemicals are produced by these plants for various processes like structure and maintenance, protection against the biotic and abiotic environmental challenges, ecological roles etc. Higher plants have a special feature of producing most and large number of phytochemicals [1]. Some chemicals among these phytochemicals have vast pharmacological properties which can be used for curing various ailments and the most important of these phytochemicals are among alkaloids, flavonoids, saponins and phenolic compounds [2, 3].

Alhagi pseudalhagi (M. Bieb.) Desv. ex B. Keller & Shap. is a perennial shrub of family Fabaceae commonly called as 'Camel thorn'. Stem has a slightly bad taste and is used as aperient, expectorant, diuretic, aphrodisiac, purifies blood; good in vomiting, small pox eruption and piles [4]. Decoctions of whole plant is cholegogic and

astringent for colitis, gastritis, stomach ulcers; to reduce water loss; for hemorrhoids & wound dressing; for dysentery, nasopharynx diseases, angina, extremity eczema and also acts as an antipyretic [5]. The entire plant is used to treat cold, fever, the damp-heat of the stomach and abdominal as well as the enteritis [6]. The plant extracts have shown good results after being tested for various cell lines for their cytotoxic activity [7]; antiulcerogenic activity [8]. Phytochemical study of Roots has already been carried [9] and chemical study of leaves has also been carried out [10]. This study was aimed to study the chemical constituents present in the Stem of *Alhagi pseudalhagi* (M. Bieb.) Desv. ex B. Keller & Shap. and active secondary metabolites present in it.

II. MATERIALS AND METHODS

The plants were collected from Gandhigram village, Akot Taluka, Akola of Vidarbha region, Maharashtra which were morphologically identified and authenticated by taxonomist Professor Dr. S.P. Rothe. The voucher specimens were deposited in the herbarium of Department of Botany, Vidyabharati Mahavidyalaya, Camp, Amravati, Maharashtra, India. The Stem of this collected plant was shade dried and then converted to powder form for further studies.

The procedures recommended in Indian Pharmacopoeia [11] and Gupta *et al.*, 2013 [12] were followed for analytical or physiochemical study.

The extraction was done by using 10 grams of plant powder and 180ml of solvent viz. Chloroform, 50% Ethanol, and Dichloromethane in the Soxhlet apparatus. The heating mantle temperature was set at evaporating temperature of the solvent and the cycles of soxhletion was done for about 6 hours. After extraction in Soxhlet apparatus for 6 hours the extracts were filtered and concentrated to 5ml using rotatory vacuum evaporator at room temperature. These concentrated samples were then used for further analysis.

The standard procedure for Qualitative analysis and quantitative analysis were followed [13]. The GC-MS analysis was carried out using gas chromatography-high resolution mass spectrometer. 2 μ l of the prepared extracts was employed for GC-MS analysis. The GC-MS analysis was carried using Alegent Hp 7880 with column of 30 meter length, with 0.25 mm internal diameter and 0.32 thickness. Helium gas was used as carrier gas at constant flow rate of 1ml/minute. Injector temperature was set at 50 °C. the Oven temperature were programmed from 50 °C to 280 °C at 10 °C/minute to 200 °C then 10 °C/ 3 minutes to 250 °C ending with a 5 minutes isothermal at 280 °C. The sample was injected in split mode as 10:80.

Interpretation on mass spectrum of GC-MS was done using the National Institute Standard and Technology (NIST) database. The mass spectrum of compounds was compared with the spectral data of known compounds present in spectral libraries (NIST). The names, molecular weight and molecular formula of the identified molecules were ascertained by this analysis.

III. RESULTS

Morphological Characters

Alhagi pseudalhagi (M. Bieb.) Desv. ex B. Keller & Shap. is a shrub of family Fabaceae commonly called as 'Camel Thorn' or 'Jawasa'. It is armed with 1-2.5 cm long spines. Leaves are drooping from the base of spines

301 | Page

or branches, smooth and simple, oblong, obtuse, glabrous, leathery, with rounded apex. Flowers are shortly pedicillate, small, 1-6 in number, borne on a spine, red papillonaceous and form panicles by auxillary racemes. Calyx is glabrous and 2-3 mm long. Corolla is Reddish and length about three times as that of Calyx. Pod is greenish gray and hard, falcate or straight, torrulose, 2.5 cm long containing 6-8 sub-reniform seeds. It has flowering period of October to March.



Figure No. 1 & 2: Habit and flower of Alhagi pseudalhagi (M. Bieb.) Desv. ex B. Keller & Shap.

Analytical Analysis

Table No 1: Analytical values of Alhagi pseudalhagi (M. Bieb.) Desv. ex B. Keller & Shap. Stem

S. No.	Parameter studied		%age value (w/w)
1.	Moisture content		8.98 ± 0.34
2.	Total Ash		7.24 ± 0.59
3.	Acid soluble ash		82.27 ± 1.75
4.	Acid insoluble ash		15.73 ± 0.88
5.	Water soluble ash		35.90 ± 0.43
6.	Water insoluble	e Ash	63.23 ± 0.37
7.	Extractive	Chloroform	4.39 ± 0.16
	values in 50% Ethanol		10.49 ± 0.46
		Dichloromethane	3.79 ± 0.36

Note: Percentage mean (n=3) \pm SD

Qualitative Phytochemical Analysis

Table No. 2 : Qualitative Phytochemical Screening of *Alhagi pseudalhagi* (M. Bieb.) Desv. ex B. Keller & Shap. Stem

S.	Constituents	Chemical tests		STEM			
No.			W	D	Ε	С	
1	Alkaloids	Wagners test	++		++	++	
		Mayers test	+ +		+ +	+ -	
2	Flavonoids	Sodium hydroxide test	++		+ +		
		Lead acetate test	++		+ +		
3	Glycosides	Killer killiani test					
		Fehlings test					
4	Phenols	Phenols test					
5	Saponins	Froathing / Foam Test	++		+ +		
6	Steroids	Salkowaski test			+ +		
		LB test			+ +		
7	Tannins	Ferric chloride test			+ +		
8	Terpenoids	Salkowaski test			+ +		

Note '+'= Present and '-'= Absent

Where, W= Water; D= Dichloromethane; E= 50% Ethanol; C= Chloroform respectively.

Quantitative Phytochemical analysis

Table No. 3: Quantitative Phytochemical analysis of *Alhagi pseudalhagi* (M. Bieb.) Desv. ex B. Keller & Shap. Stem

S. No.	Phytochemicals	(g/100 gms of dry sample)
1	Flavonoids	6.99 ± 0.72
2	Alkaloids	8.36 ± 1.33
3	Saponins	3.69 ± 0.90
4	Phenols	2.05 ± 0.72

Note: Percentage mean $(n=3) \pm SD$

Phenols: expressed as gallic acid equivalents (GAE/g) $% \left(\frac{1}{2} \right) = \frac{1}{2} \left(\frac{1}{2} \right) \left(\frac$

Flavonoids: expressed as quercetin equivalent (mg/g)

Gas Chromatography- Mass Spectrometric Analysis

Figure 3: Chromatogram of Chloroform extract of *Alhagi pseudalhagi* (M. Bieb.) Desv. ex B. Keller & Shap. Stem

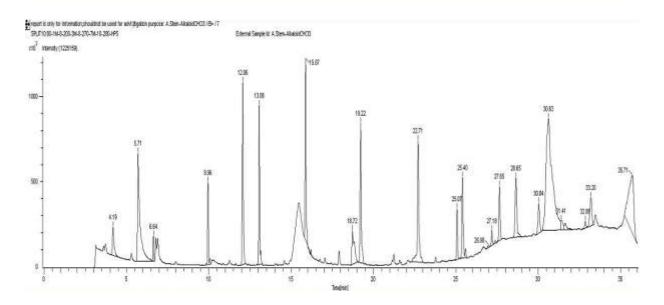


Table No. 4: Compounds identified in the Chloroform extract of Alhagi pseudalhagi (M. Bieb.) Desv. ex B.
Keller & Shap. Stem

S.No	RT	Name of compound	Peak Area %	MW	MF
1.	4.19	2-Pentenoic acid, 2-methoxy-4-methyl, methyl ester	1.66	158	C ₈ H ₁₄ O ₃
2.	5.71	Butanedioc acid, hydroxyl, dimethyl ester	9.26	162	C ₆ H ₁₀ O ₅
3.	6.64	1-Dodecene	0.74	168	C ₁₂ H ₂₄
4.	6.79	2-Propoxy-succinic acid, dimethyl ester	0.56	204	$C_9H_{16}O_5$
5.	9.96	1-Tetradecene	2.78	196	$C_{14}H_{28}$
6.	12.06	Phenol,2,4-bis(1,1-diethylethyl)-	6.97	206	C ₁₄ H ₂₂ O
7.	13.06	1-Tetradecene	5.47	196	$C_{14}H_{28}$
8.	15.45	3-O-Methyl-d-glucose	15.67	194	$C_7H_{14}O_6$
9.	15.87	1-Nonadecene	5.99	266	C ₁₉ H ₃₈
10.	19.22	1-Nonadecene	6.03	266	C ₁₉ H ₃₈
11.	22.71	1-Nonadecene	6.30	266	C ₁₉ H ₃₈
12	25.07	Stigmasterol	1.77	412	C ₂₉ H ₄₈ O
13.	25.40	1-Docosene	3.65	308	$C_{22}H_{44}$
14.	25.60	S-indacene, 1,2,3,5,6,7- hexahydro-1,1,5,5-	0.22	354	$C_{26}H_{42}$
		tetramethyl-4,8-bis (3-methylbutyl)-			
15.	27.65	Octatriacontyl pentafluropropoinate	2.24	696	$C_{41}H_{77}F_5O_2$
17.	33.20	5α–Pregnane-12,20-dione	2.05	316	$C_{21}H_{32}O_2$

(MW= Molecular Structure; MF= Molecular Formula)

Figure No. 4: Chromatogram of Dichloromethane extract of Alhagi pseudalhagi (M. Bieb.) Desv. ex B. Keller

& Shap. Stem seiSumpielit ICM A.Sten / El+ Internal Sample 1d. sp@10,00-104.6-200-304.0-200-304-00-200-HP1 this data shouldn't be used for advt, evidence or ingetion Experiment Date/Tone: 10202016140-499M Acq Detallioner Ethintoria Internsity (22110432) 138.94 4055 214 37N32 35 3.87 25,16 30.03 30 2 3 15 2 4) Telefinini

Table No. 5: Compounds identified in the Dichloromethane extract of Alhagi pseudalhagi (M. Bieb.) Desv. ex B. Keller & Shap. Stem.

S. No.	RT	Name of compound	Peak Area %	MW	MF
1.	19.54	2(4H)-Benzofuranone,5,6,7,7a-terahydro- 4,4,7a-trimethyl-(R)-	0.22	180	$C_{11}H_{16}O_2$
2.	23.95	Tetradecanoic acid	0.62	228	$C_{14}H_{28}O_2$
3.	25.14	3,7,11,15-Tetramethyl-2-hexadecen-1-ol	3.80	296	C ₂₀ H ₄₀ O
4.	27.70	n-Hexadecanoic acid	12.43	256	C ₁₆ H ₃₂ O ₂
5.	31.86	Octadecanoic acid	3.21	284	$C_{18}H_{36}O_2$
6.	35.98	3.3'-Dimethyl-1'-hydroxy-5,8-dimethoxy-2,2'- binapthalene-1,4,5',8'-tetrone	9.26	418	C ₂₄ H ₁₈ O ₇
7.	36.92	Vitamin E	20.29	430	$C_{29}H_{50}O_2$
8.	41.33	Heptacosane	2.30	380	C ₂₇ H ₅₆

305 | Page

9. 42.	.93 Y-Sitosterol	3.96	6 414	C ₂₉ H ₅₀ O
--------	------------------	------	-------	-----------------------------------

Figure No. 3: Chromatogram of 50% Ethanol extract of *Alhagi pseudalhagi* (M. Bieb.) Desv. ex B. Keller & Shap. Stem

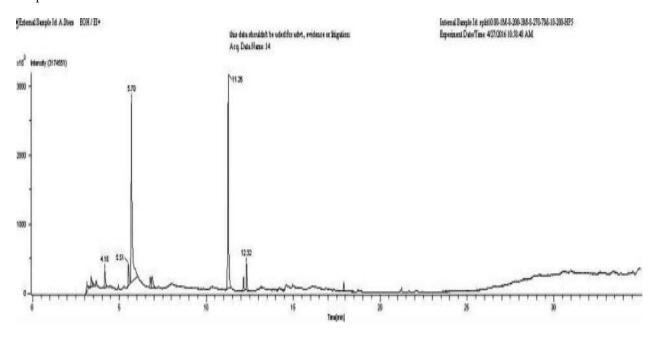


Table No. 6: Compounds identified in the 50% Ethanol extract of *Alhagi pseudalhagi* (M. Bieb.) Desv. ex B. Keller & Shap. Stem.

S. No.	RT	Name of compound	Peak Area %	MW	MF
1.	4.16	Butanedioic acid, dimethyl ester	3.37	146	$C_{6}H_{10}O_{4}$
2.	5.51	Levoglucosenone	4.44	126	C ₆ H ₆ O ₃
3.	5.70	3-Acetoxy-3-hydroxypropionic acid, methyl ester	40.04	162	C ₆ H ₁₀ O ₅
4.	6.78	2-Propoxy-succinic acid, dimethyl ester	1.87	204	$C_9H_{16}O_5$
5.	6.89	2-Propoxy-succinic acid, dimethyl ester	3.79	204	$C_9H_{16}O_5$
6.	11.26	Citric acid, trimethyl ester	34.05	234	$C_9H_{14}O_7$
7.	12.32	Trimethyl citrate	3.99	234	$C_9H_{14}O_7$

IV. DISSCUSSION

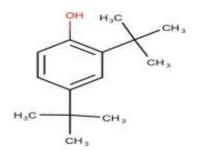
The present study was aimed to investigate the above said phytochemicals by using different solvents with different polarities for plant *Alhagi pseudalhagi* (M. Bieb.) Desv. ex B. Keller & Shap. (Fig.1 & 2). *A. pseudoalhagi* was divided into three parts viz., Roots, Stem, and Leaves. Stem was selected for the present study which was extracted in four solvents viz., Water, Dichloromethane, 50 % Ethanol and Chloroform for qualitative analysis. The quantitative analysis of Flavonoids, Alkaloids, Saponins and Phenols was done in the present study for the stem of *A. pseudoalhagi*. Three different solvents Chloroform, Dichloromethane and 50 % Ethanol were used for GC-MS analysis. GC-MS results of *A. pseudoalhagi* Stem in Chloroform are depicted in Fig. 4.4.4 and Table 4.4.12; Dichloromethane (Fig. 4.4.5 and Table 4.4.13), and 50% Ethanol (Fig. 4.4.6 and Table 4.4.14). The identified compounds were 17, 9 and 7 in solvents Chloroform, Dichloromethane and 50 % Ethanol respectively. Secondary metabolites have been found to be of utmost importance as they have vast biological importance. Among the identified compounds in the stem of *A. pseudoalhagi* one is phenol; one is terpenoid; two are Flavonoids; three are steroids while as the other identified compounds are hydrocarbons, Fattyacids, methyl esters etc.

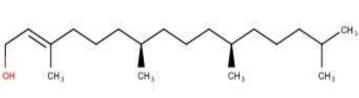
Table No. 7: Biologically active compounds present in Alhagi pseudalhagi (M. Bieb.) Desv. ex B. Keller &
Shap. Stem with their biological activities reported

S.	Name of Compound	Category	Biological activity
No.			
1.	Phenol,2,4-bis (1,1-	Phenol	Anti-bacterial activity [14]; Antifungal activity and
	dimethylethyl)-		Antioxidant [15]; Anti-inflammatory activity [16];
			Anticancerous [17].
2.	3,7,11,15-Tetramethyl-2-	Terpenoid/	Anti-inflammatory [18]; Antimicrobial, diuretic [19];
	hexadecen-1-ol (Phytol)	Diterpene	Anti-cancer [20]; Cancer preventive [21]; Joint
			dislocation, Hernia and Antimalarial [22].
3.	2(4H)-Benzofuranone,5,6,7,	Coumaran /	Antihelminthic, Anti-inflammatory, Antidiarrhoeal [23].
	7a-terahydro-4,4,7a-	Flavonoid	
	trimethyl-(R)-		
4.	Y-Sitosterol	Steroid	Anticancerous [24]; Anti breast cancer [25];
			Antioxidative [26]; Anti-inflammatory [27]; Uterotonic,
			Estrogenic [28].
5.	5α–Pregnane-12,20-dione	Steroid	Anticancerous [29, 30]; Antibacterial activity [31].

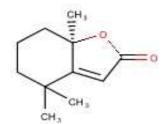
6.	Stigmasterol	Steroid	Antioxidant & reduce blood cholesterol level [32]; Anti-
			osteoarthitic [33]; Cyto-toxicity activity [34]; Anti-tumor
			activity [35]; anti-HIV reverse transcriptase [36];
			Thyroid inhibiting properties, Precursor of progesterone,
			Antimicrobial, Anti-asthama, Anti inflammatory,
			Diuretic [22]; inhibit Na ⁺ K ⁺ pump ATPases and
			influence prostrate metabolism and growth [37].
7.	3,3'-Dimethyl-1'-hydroxy-	Flavonoid/	Anti-cancer activity [38]
	5,8-dimethoxy-2,2'-	Napthoquinone	
	binapthalene-1,4,5',8'-	1 1	
	tetrone (Diospyrin)		

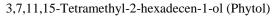
Molecular structures of the biologically active compounds from Alhagi pseudalhagi Stem

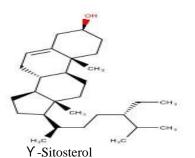




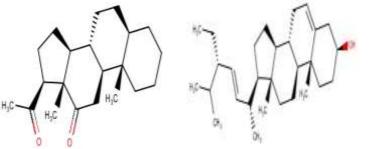
Phenol,2,4-bis (1,1-dimethylethyl)-

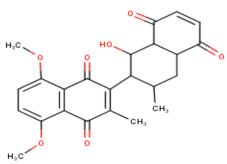






2(4H)-Benzofuranone, 5,6,7,7a-terahydro-4,4,7a-trimethyl-(R)-





308 | Page

5α–Pregnane-12, 20-dione

Stigmasterol

3,3'-Dimethyl-1'-hydroxy-5,8-dimethoxy- 2,2'binapthalene-1,4,5',8'-tetrone (Diospyrin)

V. CONCLUSION

The results suggest that this plant has vast variety of phytochemicals which can be used as effective remedy for various ailments and drug formulations in future either alone or in combination with other suitable agents. The ethnic claims of this plant were also verified by the present study as there is presence of phytochemicals which are responsible to show these activities.

VI. ACKNOWLEDGEMENTS

The authors express sincere thanks to Sophisticated Analytical Instrument Facility (SAIF), Indian Institute of Technology (IIT), Mumbai for supporting instrumental analysis for this research work. The authors are also thankful to Dr. V.R. Deshmukh, Head Department of Botany, VBMV, Camp, Amravati for providing necessary laboratory facilities.

REFERENCES

- [1] M.C. Castello, A. Phattak, N. Chandra, and M. Sharon, Antimicrobial activity of crude extracts from plant parts and corresponding calli of *Bixa orellana* L., *Indian J. Exp. Biol.* 40, 2002, 1378-1381.
- [2] A.F. Hill, Economic Botany (McGraw-Hill Book Company, INC., Tokyo, 1952)
- [3] K.H. Pandey, P. Khadka, S.K. Thapa, and S. Panta, Phytochemical screening and Analysis of Antioxidant Activity of *Crateva unilocularis* Buch,-Ham. Leaf, *International Journal of Pharmaceutical Research Scholars*, 2(4), 2013, 123-130.
- [4] B. Yadav, A Flavanone from Alhagi pseudalhagi, J. Biol. Chem. Research, 31(2), 2014, 1249-1255.
- [5] A. Sultan, M. Mohammadnor, and K.A. Eshbakova, Chemical constituents of *Alhagi pseudoalhagi.*, *Chemistry of Natural Compounds*, 47(1), 2011, 140-141.
- [6] G. J. Zhang, The Chemical Composition Of Alhagi Research, Master Thesis, Natural medicinal chemistry, China, 2010.
- [7] Behzad Sahar, Atefeh Pirani and Mahmoud Mosaddegha, Cytotoxic Activity of Some Medicinal Plants from Hamedan District of Iran, *Iranian Journal of Pharmaceutical Research*, *13*, 2014, 199-205.
- [8] M.K. Gharib Naseri, and Seyyed Ali Mard, Gastroprotective effect of *Alhagi maurorum* on exprimental gastric ulcer in rats, *Pak J Med Sci.*, 23(4), 2007, 570-573.
- [9] N.A. Wagay and S.P. Rothe, Investigations on secondary metabolites of *Alhagi pseudalhagi* (M. Bieb.) Desv ex B. Keller & Shap. Leaves using GC-MS, *Journal of Pharmacognosy and Phytochemistry*, 5(5), 2016, 114-118.

- [10] N.A. Wagay, Sheikh Ummar, S.P. Rothe and A.A. Maheshwari, Chemoprofiling of *Alhagi pseudalhagi* (M. Bieb.) Desv. ex B. Keller & Shap. Roots, *International Journal of Applied Research*, 3(3S), part E, 2017, 147-151.
- [11] Anonymous, *The Indian Pharamacopoeia* (2nd Edition, Goverment of India publication, New Delhi, 1966).
- [12] A. Gupta, A. Joshi, and B. Dubey, Comparative pharmacognostical and phytochemical evaluation of two species of *Cyathocline, International Journal of Biomedical Research*, 04(10), 2013, 538-545.
- [13] P. V. Pulate, N.A.Wagay and V. R. Deshmukh, Phytochemical, Ethnomedicinal and Anatomical study of *Canthium parviflorum, World Journal of Pharmacy and Pharmaceutical Sciences, 4(11), 2015, 1464-1482.*
- [14] K.M. Kuppuswamy, J. Bhavana, and A. Sumathy, GC-MS analysis of chloroform extract of *Croton bonplandianum*, Int J Pharm Bio Sci., 4(4), 2013, (P)613-617.
- [15] N. Raja Rajeswari, S. RamaLakshmi, and K. Muthuchelian, GC-MS Analysis of bioactive components from the ethanolic leaf extract of *Canthium dicoccum* (Gaertn.) Teijsm & Binn., *J. Chem. Pharm. Res.*, 3(3), 2011, 792-798.
- [16] Sanitha Phillips, Mudiganti Ram Krishna Rao, K. Prabhu, Minu Priya, S. Kalaivani, Aparna Ravi and Shruti Dinakar, Preliminary GC-MS analysis of an Ayurvedic medicine "Kulathadi Kashayam", J. Chem. Pharm. Res., 7(9), 2015, 393-400.
- [17] D.M. Pereira, P. Patrícia Valentao, J.A. Pereira, and P.B. Andrade, Phenolics: From Chemistry to Biology, *Molecules*, 14, 2009, 2202-2211.
- [18] M. Ogunlesi, W. Okiei, E. Ofor, and A.E. Osibote, Analysis of the essential oil from the dried leaves of *Euphorbia hirta* Linn. (Euphorbiaceae), a potential medication for asthma, *Afric. J. Biotech.*, 8, 2009, 7042-7050.
- [19] G. Rajeswari, M. Murugan, and V.R. Mohan, GC-MS analysis of bioactive components of *Hugonia mystax L*. (Linaceae), *RJPBCS*, 3(4), 2012, 301-308.
- [20] M. Himaja and Moonjit Das, Phytochemical screening, GC-MS analysis and biological activities of *Ipomoea eriocarpa* leaf extracts, *Int J Pharm Sci.*, 6(4), 2014, 592-594.
- [21] M. Sermakkani and V. Thangapandian, GC-MS analysis of *Cassia italica* leaf Methanol Extract, *Asian Journal of Pharmaceutical and Clinical Research*, *5*(2), 2012, 90-94.
- [22] R. Dandekar, B. Fegade and V.H. Bhaskar, GC-MS analysis of phytoconstituents in alcohol extract of Epiphyllum oxypetalum leaves, Journal of Pharmacognosy and Phytochemistry, 4(1), 2015, 149-154.
- [23] Vijisaral Elezabeth D. and Subramanian Arumugam, GC MS Analysis of Ethanol Extract of *Cyperus rotundus* Leaves, *Int.J.Curr.Biotechnol.*, 2(1), 2014, 19-23.
- [24] C. Scholtysek, A.A. Krukiewicz, J.L. Alonso, K.P. Sharma, P.C. Sharma, and W.H. Goldmann, Characterizing components of the Saw Palmetto Berry Extract (SPBE) on prostate cancer cell growth and traction, *Biochem Biophys Res. Commun.*, 379 (3), 2009, 795-798.
- [25] Bruce J. Grattan Jr., Plant Sterols as Anticancer Nutrients: Evidence for Their Role in Breast Cancer, *Nutrients*, 5, 2013, 359-387.

- [26] T. Wang, K.B. Hicks, and R. Moreau, Antioxidant activity of phytosterols, oryzanol, and other phytosterol conjugates, J. Am. Oil Chem. Soc., 79, 2002, 1201–1206.
- [27] Sosa Azhar Abduameer, Suhaila Husaein Bagi, and Imad Hadi Hameed, Analysis of bioactive chemical compounds of *Euphorbia lathyrus* using gas chromatography-mass spectrometry and Fourier-transform infrared spectroscopy, *J. Pharmacognosy Phytother*, 8(5), 2016, 109-126.
- [28] James A. Duke, Mary Jo B. Godwin, J. DuCdlier, Peggy-Ann K. Duke, *Handbook of Medicinal Herbs* (2nd edition, CRC press, USA, 2002).
- [29] P.G. Bradford, and A.B. Awad, Phytosterols as anticancer compounds, *Molecular Nutrition and Food Research*, 51(2), 2007, 161–170.
- [30] T.A. Woyengo, V.R. Ramprasath, and P.J.H. Jones, Anticancer effects of phytosterols, *European Journal* of Clinical Nutrition, 63(7), 2009, 813–820.
- [31] R.F. Epand, P.B. Savage, and R.M. Epand, Bacterial lipid composition and the antimicrobial efficacy of cationic steroid compounds (Ceragenins), *Biochem Biophys Acta*, *1768* (*10*), 2007, 2500-2509.
- [32] J. Zawistowaski, Tangible Health Benefits of Phytosterol: Functional Foods. In: J. Smith and E. Charter (eds.), *Functional Food Product Development* (Wiley Blackwell. 2010, 362-372.)
- [33] O. Gabay, C. Sanchez, C. Salvat, F. Chevy, M. Breton, and G. Nourissat, Stigmasterol: a phytosterol with potential anti-osteoarthritic properties, *Osteoarthritis Cartilage*, *18*(1), 2010, 106-116.
- [34] J.G. Huang, L.J. Zhou, H.H. Xu and W.O. Li, Insecticidal and cytotoxic activities of extracts of *Cacalia tangutica* and its two active ingredients against *Musa domestica* and *Aedes albapictus*, *Journal of Economic Entomology*, 102 (4), 2009, 1444-1447.
- [35] Y. Kasahara, K. Kumaki, S. Katagiri, K. Yasukawa, S. Yamanouchi, and M. Takido, *Carthami flos* extract and its component, stigmasterol, inhibit tumour promotion in mouse skin two stage carcinogenesis, *Phytotherapy Research*, 8(6), 1994, 327-331.
- [36] G.G. De Oliveira, J. A. De S. Pereira junior, I.V.G.A. Bastos, R.C.O. Sampaio Filh, N.P. Lopes, and S.J. De melo, Phytochemical Investigation of Chloroform extract from Root, Stem and Leaf of Adenocalymma imperatoris-maximilianii (Wawra) L.G. Lohmam (Bignoniaceae), Int J Pharm Bio Sci., 5(3), 2014, 70-78.
- [37] E. Bombardelli and P. Morazzoni, Prunus africana (Hook.f.) Kalkm, Fitoterapia, 68, 205-218.
- [38] S. Sagar, M. Kaur, K.P. Minneman and V.B. Bajic, Anti-cancer activities of diospyrin, its derivatives and analogues, *Eur J Med Chem.*, 45(9), 2010, 3519-30.