

PAPER TITLE: ISOLATION AND IDENTIFICATION OF PHOSPHATE SOLUBILIZING FUNGI FROM RHIZOSPHERE SOIL OF CHICKPEA PLANT

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ABSTRACT

The present investigation was carried out for understanding and exploring the indigenous phosphate solubilizing fungi and their efficient utilization as a potential biofertilizer for the improvement of soil fertility. In this study total of 23 fungi were isolated from rhizospheric soil of chickpea plants. Out of 23, thirteen fungi were showed as phosphate solubilizers on the Pikovskaya's Agar medium. Fungi that showed halo zones around the colonies are good phosphate solubilizers and belong mainly to the genera of *Aspergillus*, *Penicillium*. The present study would help in utilizing the indigenous phosphate solubilizing fungal strains as an effective biofertilizers by the farmers.

Keywords: *Aspergillus*, *Biofertilizer*, *Penicillium*, *Phosphate solubilizing fungi* & *Solubilization*

1.INTRODUCTION

Phosphorus is one the most essential elements for plant growth after nitrogen. However, the availability of this nutrient for plants is limited by different chemical reactions especially in arid and semi-arid soils. Phosphorus plays a significant role in several physiological and biochemical plant activities like photosynthesis, transformation of sugar to starch, and transporting of the genetic traits.

Phosphate solubilizing microorganisms refer to a group of soil microorganisms that as components of phosphorus cycle, can release it from insoluble sources by different mechanisms [1]. Phosphate solubilizing fungi and bacteria are known as effective organisms in this process [2]. Fungi are the important components of soil microbes typically constituting more of the soil biomass than bacteria, depending on soil depth and nutrient conditions. Fungi have been reported to have greater ability to solubilize insoluble phosphate than bacteria [3]. A wide range of soil fungi are reported to solubilize insoluble phosphorous such as *Aspergillus niger* and *Penicillium sp.* which are the most common fungi capable of phosphate solubilization [4]. Exploration of phosphate solubilizing microorganisms has been conducted by many researchers from soils, mangrove and rhizosphere (5). Since large population of Chhattisgarh state is dependent on agriculture the present investigation is aimed to isolate some fungal strains that may have high efficiency for phosphate solubilization.

II.MATERIAL AND METHOD

The present investigation was carried out in the SoS Life Sciences, Pt. Ravishankar Shukla University, Raipur, Chhattisgarh.

2.1 Study Site

Raipur is situated in East Central part of Chhattisgarh at latitude of 21⁰16' N, longitude 81⁰36' E and altitude 289.5 m above mean sea level. The climate of Raipur is falling under sub-humid with mean annual rainfall of about 1489 mm out of which 90 per cent (1348 mm) is received during monsoon (June to September). During *rabi*, (December to February) only 8 mm of rainfall is being received on an average. The maximum temperature ranged from 24.4 to 42.6 °C and minimum temperature ranged between 10.0 to 27.5 °C respectively.

2.2 Collection of soil samples

Soil samples were collected from rhizosphere of Chickpea plantation from 3 different villages of Raipur city of Chhattisgarh state. Samples were collected in polythene bags, transported to laboratory and stored in refrigerator for further processing. Soil samples were separated from roots, air dried at room temperature, crushed, sieved and collected in separate polythene bags. pH of the samples was recorded using pH meter (Elico made).

2.3 Culture media for isolation

Pikovskaya's (6) agar medium (HIMEDIA) was used for the isolation and maintenance of phosphate solubilizing fungi. It contained (g litre⁻¹) Dextrose 10; Calcium phosphate 5; Ammonium sulphate 0.5; Potassium chloride 0.2; Magnesium sulphate 0.1; Manganese sulphate 0.0001; Yeast extract 0.5; Ferrous sulphate 0.0001, Agar 15. The pH of medium was 7.0 (± 0.2).

Potato dextrose agar (PDA, HIMEDIA) was used for the isolation maintenance of fungal cultures. It contained (g.litre⁻¹) potato infusion 200; Dextrose 20; Agar 15 and the pH of medium was 5.6 (± 0.2) The pH of culture media was adjusted using 1N NaOH or 1N HCl. Media were sterilized by autoclaving at 121 °C for 15 min.

2.4 Isolation

Suitable dilutions prepared in 0.85% saline were spread on plates containing Potato Dextrose Agar (PDA) medium. The plates were kept for incubation at 28±2°C for 5-7 days. Fungal colonies were subcultured several times on PDA plates till the appearance of pure cultures. The isolates were stored in refrigerator on PDA slants for further studies.

2.5 Screening

The isolates were screened by inoculating on plates containing Pikovskaya's Agar (PKA) medium (6) amended with 0.5% tricalcium phosphate (TCP) as insoluble phosphate source and were incubated at 28±2°C for 5 days. Fungal colonies with clear halozone around them were screened as phosphate solubilizers.

2.6 Identification

The fungal cultures were identified on the basis of colony characteristics and microscopic examination (7, 8, and 9). Some of the fungal isolates have been sent and deposited to NFCCI for identification

III.RESULTS AND DISCUSSIONS

Total 23 fungi were isolated from rhizospheric soils of chickpea plantation and 13 were screened as phosphate solubilizers based on appearance of clear halozone on Pikovskaya's agar medium (**Table**). Fungi that showed halo zones around the colony are good phosphate solubilizers and belong mainly to the genera of *Aspergillus*, and *Penicillium*. *Aspergillus niger* was found to be the dominant group followed by *Penicillium* sp. and other species of *Aspergillus*. The similar results were highlighted by Mahamuni et al. (10) and Deepa et al. (11). The higher number of the species of the fungus can be attributed to its ability to grow in diverse conditions (12). Soil fungi make a very important part of the ecosystem along with other microbes in turnover of the biomass (13). It is known that the species of *Aspergillus*, *Trichoderma*, *Penicillium*, *Fusarium*, *Mucor* etc. are the dominantly occurring fungi isolated from different rhizospheric soils and screened for phosphate solubilization by many workers (14 and 15).

IV.PHOTOGRAPH OF HALOZONE FORMATION & ASPERGILLUS NIGER



V.TABLE

S. No	Village Name (Raipur)	Soil p ^H	Isolated Fungi name	Formation of Halozone
01	Jora Village	7.70	<i>Aspergillus</i> sp.J1	Yes
			<i>Aspergillus niger</i>	Yes
			<i>Aspergillus fumigatus</i>	Yes
			<i>Alternaria alternata</i>	Yes
			<i>Penicillium</i> sp.J1	No
			<i>Aspergillus</i> sp.J2	Yes
			<i>Aspergillus</i> sp. J3	Yes
			<i>Rhizopus</i> sp.	No
			<i>Fusarium</i> sp.	No
			<i>Penicillium</i> sp. J2	No
02	Tekari	6.8	<i>Curvularia</i> sp.	No
			<i>Aspergillus</i> sp.	Yes
			<i>Fusarium</i> sp	No
			<i>Aspergillus niger</i>	Yes
			<i>Penicillium</i> sp.	Yes
			<i>Rhizopus</i> sp.	No
03	Nakti	7.2	<i>Aspergillus niger</i>	Yes
			<i>Aspergillus</i> sp.	Yes
			<i>Penicillium</i> sp.	Yes
			<i>Curvularia</i> sp.	No
			<i>Mucor</i> sp.	No
			<i>Rhizopus</i> sp.	No
			<i>Alternaria</i> sp	Yes

Table: Screening of the phosphate solublizing properties by the isolated fungi on the basis of halo zone formation.

VI.CONCLUSION

It is concluded that phosphate solubilizing fungi were present in the rhizosphere of chickpea. Further research should be continued with these efficient strains especially *Aspergillus niger* and *Penicillium* sp. These isolates may be used as bioinoculant in agriculture for increasing crop productivity.

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